

**A NOVEL MICROORGANISM ISOLATED FROM CHINESE ELM (*ULMUS*
SP.) AND PROCESS FOR PREPARING EXOPOLYSACCHARIDES BY
EMPLOYING THE MICROORGANISM**

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Background of the Invention

Field of the Invention

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The present invention relates to a novel microorganism isolated from Chinese elm(*Ulmus* sp.) and a process for preparing immunostimulating exopolysaccharides with anticancer activity by employing the microorganism, more specifically, to a novel *Enterobacter* sp. isolated from the root bark of Chinese elm(*Ulmus* sp.) alive, which produces immunostimulating exopolysaccharides with anticancer activity, a process for preparing the exopolysaccharides by culturing the microorganism, the exopolysaccharides thus prepared and novel practical uses thereof.

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Description of the Related Art

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Polysaccharides have been produced from natural products such as plants and seaweeds, or microorganisms and widely used for the materials for foodstuffs, medicine, etc. In light of the various uses of polysaccharides, intensive research has been actively undertaken to produce polysaccharides with novel structure and sugar composition, and to explore their potential uses.

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Recently, it has been reported that polysaccharides existing in exterior of cell wall play an important role as a messenger for signal transduction between cells, which arouse great interests in polysaccharides. Examples of commercialized exopolysaccharides produced by microorganisms include xanthan gum, pullulan, gellan gum, curdlan, hyaluronic acid, etc.

Meanwhile, in Korea and China, Chinese elm (elm tree as common name) had been used for a long time as a therapeutic agent, as well as for relieving famine. The

root bark of Chinese elm has been used in a dried form for oriental medicine or folk remedy, and its pharmacological properties are described in ancient literatures of oriental medicine such as Bonchogangmok, Euhakibmoon, Hyangyakjibsongbang, and Dongeubogam. According to such medical literatures, the root bark of Chinese elm is known to be a powerful remedy for a rash and an abscess, effective on gastric ptosis, gastric ulcer, and duodenal ulcer, and also working on a mild stomachache, a digestive disorder, urinating disorder, empyema, otitis media, an abscess, a festered wound such as a swelling and a boil, skin conditioning, and uterine- and breast-associated diseases.

In this connection, the present inventors also found that water-soluble polysaccharides(proteoglycans) extracted from the root bark of Chinese elm have an anticancer activity by immunostimulation.

However, it has been revealed that water-soluble polysaccharides(proteoglycans) extracted from the root bark of Chinese elm may bring about destruction of the plant species and their qualities cannot be controlled in an efficient manner owing to different compositions depending on the area and timing of taking the plants.

Under the circumstances, there are strong reasons for exploring and developing alternative process for preparing the exopolysaccharides of uniform quality in a massive manner.

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Summary of the Invention

The present inventors have made an effort to solve the problems mentioned above, and isolated a novel microorganism of *Enterobacter* sp. which produces exopolysaccharides with immunostimulating activity, from the root bark of Chinese elm. Furthermore, they found that: the exopolysaccharides produced by fermentation of the said microorganism possess an anticancer activity by immunostimulation which is similar to physiological activity of the water-soluble polysaccharides(proteoglycans) extracted from the root bark of Chinese elm; and, their mass production can be

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accomplished with a uniform quality by controlling medium and culture conditions, without destruction of the plant species.

One aspect of the present invention is to provide an isolated microorganism identified by accession number KCTC 0687BP.

5 Another aspect of the present invention provides a method of producing an exopolysaccharide. The method comprises the steps of providing an isolated microorganism identified by accession number KCTC 0687BP, and culturing the microorganism in a medium so as to allow production of an exopolysaccharide. The method further comprises isolating the exopolysaccharide from a mixture comprising
10 the culture medium, the microorganism and the exopolysaccharide. The culture medium comprises a carbon source selected from the group consisting of glucose, sucrose, fructose, rhamnose, galactose, arabinose, mannitol, lactose, gluconate, xylose and mixtures thereof. The culturing is performed at a temperature ranged from about 25 °C to about 38 °C under aeration at a flow rate ranged from about 0.1 vvm to about 1.5
15 vvm and under agitation at an agitation speed ranged from about 150 to about 500 rpm. The isolation of the exopolysaccharide comprises: removing cells from the culture mixture; and dialyzing a resulting mixture so as to isolate the exopolysaccharide. The isolation further comprises lyophilizing the separated exopolysaccharide. The removal of cells comprises: centrifuging the culture mixture to obtain a supernatant; precipitating
20 a mixture comprising the exopolysaccharide; dissolving the precipitate in a liquid; and removing remaining cells. The present invention further provides a composition obtainable by the method.

Another aspect of the present invention provides a composition comprising an isolated exopolysaccharide from an *Enterobacter* species, wherein the species is
25 obtained from root bark of Chinese elm, *Ulmus* species and the exopolysaccharide has a molecular weight ranged from about 100,000 to about 1,000,000. The isolated exopolysaccharide comprises sugar in an amount ranged from about 40 wt.% to about 75 wt.%. The isolated exopolysaccharide comprises acidic sugar in an amount ranged from about 5 wt.% to about 15 wt.%. The isolated exopolysaccharide comprises protein

in an amount ranged from about 10 wt.% to about 25 wt.%. The isolated exopolysaccharide comprises glucose, fructose, galactose, fucose and glucuronic acid. The isolated exopolysaccharide comprises 10-30 wt.% glucose, less than 1 wt.% fructose, 10-15 wt.% galactose, 8-12 wt.% fucose and 40-70 wt.% glucuronic acid.

5 Still another aspect of the present invention provides a method of inducing immune cell proliferation, which comprises providing cells and contacting the exopolysaccharide with the cells, thereby stimulating proliferation of immune cells. The method further comprises identifying immune cells in need of an induction of proliferation and/or measuring immune cell proliferation.

10 Still another aspect of the present invention provides a method of inhibiting proliferation of cancer cells. The method comprises providing a cancer cell and contacting the exopolysaccharide with the cancer cell. The cancer cells comprising melanoma cells.

15 Still further aspect of the present invention provides a method of inhibiting cancer cell proliferation in a mammal. The method comprises identifying a mammal in need of an agent that inhibit cancer cell proliferation and providing the mammal with the exopolysaccharide.

Brief Description of the Drawings

20 The above, the other objects and features of the invention will become apparent from the following descriptions given in conjunction with the accompanying drawings, in which:

Figure 1 is a schematic representation of the process for isolating a microorganism from the root bark of Chinese elm.

25 Figure 2 is a schematic representation of the process for preparing the exopolysaccharide by fermentation of a novel microorganism.

Figure 3 is a graph showing a profile of batch fermentation of the microorganism producing exopolysaccharide.

25-38 °C, under an agitation speed of 150-500rpm, pH 4.0-7.5 and an aeration flow rate of 0.1-1.5vvm to produce exopolysaccharides, which was subsequently isolated and purified, preferably, followed by lyophilization. The culture medium comprises a carbon source such as glucose, sucrose, fructose, rhamnose, galactose, arabinose, mannitol, lactose, gluconate, xylose, and mixture thereof.

For the preparation of the exopolysaccharides, the microorganism is cultured in a medium to produce exopolysaccharides, and then, the exopolysaccharides are isolated from the culture, which comprises the steps of: centrifuging the culture to obtain a supernatant; adding an organic solvent such as acetone, ethanol, methanol or propanol to the supernatant at a ratio of 1:1-1:5(v/v), preferably, a ratio of 1:2-1:3(v/v), to give a precipitate; dissolving the precipitate in distilled water and removing remaining cells by filtration or centrifugation; and, dialyzing the resultant against a membrane with 4,000-14,000 MWCO to discard residual saccharides and low molecular weight materials. After dialysis, filtration or centrifugation may be further carried out to remove remaining cells, if necessary.

Exopolysaccharides thus produced have a molecular weight of 100,000 to 1,000,000 and consist of 40-75% of total sugar, 5-15% of total acidic sugar and 10-25% of total protein. Compositions and contents of sugars in the exopolysaccharides are 10-30% glucose, less than 1% fructose, 10-15% galactose, 8-12% fucose and 40-70% glucuronic acid.

In the course of isolating exopolysaccharides from the culture, biological process employing enzymes such as trypsin, pepsin and papain, or chemical process employing chemicals such as trichloroacetic acid may be additionally applied to remove protein impurities in the exopolysaccharides.

Changes in intrinsic viscosity of the exopolysaccharide depending on concentration and pH have been studied, and a comparison of rheological properties of the exopolysaccharide with those of xanthan gum has been made to find out the exopolysaccharide of the invention has similar pseudoplastic rheological properties to commercially available xanthan gum.

Further, the investigations for anticancer activity by immunostimulation, employing exopolysaccharides produced by the said fermentation, were intended to propose their uses possible on the basis of their experimental results. Anticancer activity by immunostimulation of the exopolysaccharide has been determined by immunostimulating assays *in vitro* (immune cell proliferation, mixed lymphocyte reaction and direct mitogenicity for immune cells), and by immunostimulating assays *in vivo* (test of acute toxicity in a septic shock model system, and anticancer activity assay in a B16 melanoma model system).

In these experiments, the exopolysaccharides exhibited high immunostimulating activities in assays for immune cell proliferation, mixed lymphocyte reaction, and direct mitogenicity, moreover, survival rate of melanoma mice increased by 138.1% at a dose of 0.3mg/kg in assays for anticancer activity *in vivo*, by virtue of immunostimulation. On the other hand, it has been observed and reported that the anticancer activity of the water-soluble polysaccharides (proteoglycans) extracted from root bark of Chinese elm increases by 139.2% in survival rate of melanoma mice at a dose of 3mg/kg. Thus, it was found that with a 1/10 concentration the immunostimulating anticancer activity of the exopolysaccharide derived from the microorganism of the invention was similar to those of the said water-soluble polysaccharides (proteoglycans) extracted from the root bark of Chinese elm.

Therefore, the present invention provides practical uses of the exopolysaccharides for anticancer agents, immunoenhancers and foodstuffs as an active ingredient. Besides such specific uses, the exopolysaccharide of the invention may be used for general purposes of polysaccharides such as emulsifier, stabilizer, adhesive, coagulant, agglutinating agent, lubricant, film-forming agent, thickener, suspending agent, and other industrially important polymer mediators.

Anticancer agents or immunostimulants comprising the exopolysaccharides of the invention as an active ingredient may be subjected to enteral (oral or inhaled) or parenteral (nonoral) (e.g., intravenous, subcutaneous, transcutaneous and rectal) administration, and the said agents may be prepared depending on purposes, in various

preparations such as tablets, capsules, granules, powders, suppositories, ointments, injections, emulsions, suspensions, and syrups. The said various formulations may be prepared by conventional methods using nontoxic additives such as excipient, binder, disintegrator, lubricant, preservative, antioxidant, isotonic agent, buffer, coating agent, sweetener, solubilizer, base, dispersant, stabilizer, coloring agent, etc. Examples of nontoxic additives available are listed below.

First, excipients include starch and its derivatives (dextrin, carboxymethylstarch, etc.), cellulose and its derivatives (methylcellulose, hydroxypropylmethylcellulose, etc.), saccharides (lactose, sucrose, glucose, etc.), silicic acid and silicates (natural aluminium silicate, magnesium silicate, etc.), carbonates (calcium carbonate, magnesium carbonate, sodium bicarbonate, etc), aluminium hydroxide, magnesium, synthetic hydrotalcite, polyoxyethylene derivatives, glycerine monostearic acid, sorbitan monooleic acid, etc.

Binders include starch and its derivatives (α -starch, dextrin, etc.), cellulose and its derivatives (ethyl cellulose, sodium carboxymethyl cellulose, hydroxypropylmethyl cellulose, etc.), gum arabic, gum tragacanth, gelatin, saccharides (glucose, sucrose, etc.), ethanol, polyvinylalcohol, etc.

Disintegrators include starch and its derivatives (carboxymethyl starch, hydroxypropylmethyl starch, etc.), cellulose and its derivatives (sodium carboxymethyl cellulose, hydroxypropylmethyl cellulose, etc.), carbonates (calcium carbonate, calcium hydrogen carbonate, etc.), gum tragacanth, gelatin, sugars, etc.

Lubricants include stearic acid, calcium stearate, magnesium stearate, talc, silicic acid and silicates (rigid silica, natural aluminium silicate, etc.), titanium oxide, calcium hydrogen phosphate, dried aluminium hydroxide gel, etc.

Preservatives include *p*-hydroxybenzoate, sulfites (sodium sulphite, sodium pyrosulfites, etc.), phosphates(sodium phosphate, poly (calcium phosphate), poly (sodium phosphate), sodium metaphosphate, etc.), alcohols (chlorobutanol, benzylalcohol, etc.), benzalconium chloride, benzetonium chloride, phenol, cresol,

cresol chloride, dihydroacetic acid, sodium dihydroacetate, glycerin sorbic acid, sugars, etc.

5 Antioxidants include sulfites (sodium sulfite, sodium hydrogen sulfite, etc.), rongalite, erythorbic acid, L-ascorbic acid, cysteine, thioglycerol, butyl hydroxy anisol, dibuthylhydroxy toluene, propylgallic acid, ascorbylpalmitate, dl- α -tocopherol, etc.

Isotonic agents include sodium chloride, sodium nitrate, potassium nitrate, dextrin, glycerin, glucose, etc.

10 Coating agents include cellulose derivatives (hydroxypropyl cellulose, celluloseacetate phthalate, hydroxypropylmethyl cellulose phthalate, etc.), shellac, polyvinylpyrrolidon, polyvinylpyridine (poly-2-vinylpyridine, poly-2-vinyl-5-ethylpyridine, etc.), polyvinylacetyl diethylaminoacetate, polyvinylalcohol phthalate, methacrylate, methacrylate copolymer, etc.

Sweeteners include saccharides (glucose, lactose, sucrose, etc.), sodium saccharin, sugar alcohol, etc.

15 Solubilizers include ethylenediamine, nicotinamide, sodium saccharin, citric acid, citrate, sodium benzoate, soaps, polyvinylpyrrolidone, polysorbate, sorbitan, fatty acid ester, glycerin, propyleneglycol, benzylalcohol, etc.

20 Bases include lipid (lard, etc.), vegetable oil, animal fat, lanolin acid, petrolatum, paraffin, wax, resin, bentonite, glycerin, glycol oil, high alcohol (stearylalcohol, cetanol, etc.), etc.

Dispersants include gum arabic, gum tragacanth, cellulose derivatives (methyl cellulose, etc.), stearic acid polyester, sorbitan sesquioleate, aluminium monostearate, sodium alginate, polysorbate, sorbitan fatty acid ester, etc.

25 Stabilizers include sulfites (sodium hydrogen sulfite, etc.), nitrogen, carbon dioxide, etc.

In the preparation of the pharmaceutical formulations, the content of the exopolysaccharides of the invention, may differ according to the type of formulations, but in general is 0.01 to 100 wt%.

Dosage level of anticancer agents or immunostimulants of the invention may vary in a wide range depending on the kinds of warm-blood animals to be treated, including a human, the progress of disease, the decision of doctor, etc. However, the effective dosage may be determined within the range from 0.01 to 30mg/kg (based on the exopolysaccharides or their pharmaceutically acceptable salts) for an enteral administration, and within the range from 0.01 to 10mg/kg for parenteral administration, per day for a 60kg adult. The daily dose mentioned above may be administered once a day or divided into several doses, which may be optionally changed depending on the progress of the disease and doctor's decision.

The present invention is further illustrated in the following examples, which should not be taken to limit the scope of the invention.

Example 1: Isolation and identification of a novel microorganism producing exopolysaccharides from root bark of Chinese elm.

The root of Chinese elm was picked from a hill near Kimhae, Wando arboretum, and Cheongsando, Korea. After removing soil from the root, the bark was taken off, chopped, and thoroughly ground with distilled water in a blender. During this process, water-soluble polysaccharides were extracted and the solution became viscous. The viscous solution was streaked on a solid agar medium of MYGP (malt extract 3g/L, yeast extract 3g/L, glucose 10g/L, bactopectone 5g/L).

After incubating for about 2 days at 30 °C, colonies producing exopolysaccharides were isolated from various colonies on a solid agar medium. The isolated colonies were checked for production of exopolysaccharides in MYGP liquid medium (100 ml) under a condition of 30 °C and 200 rpm, and the strain with the highest yield was isolated and identified. Identification of the strain was performed at the Collection for Type Cultures (KCTC) affiliated with the Korea Research Institute of Bioscience and Biotechnology (KRIBB), an international depository authority, with various analysis methods such as fatty acid analysis (MIDI), API 20E analysis, BIOLOG analysis, and 16S rRNA sequencing. Figure 1 is a schematic representation of

the isolation process of microorganism producing exopolysaccharides from the root bark of live Chinese elm.

5 Upon overall consideration of the above-mentioned analyses, the said microorganism producing exopolysaccharides was identified as a novel strain belonging to *Enterobacter* sp. Therefore, the present inventors named the identified bacterial strain "*Enterobacter* sp. SSYL", and deposited with the Korean Collection for Type Cultures (KCTC, #52, Oun-dong, Yusong-ku, Taejon, Republic of Korea) affiliated with the Korea Research Institute of Bioscience and Biotechnology (KRIBB), an international depository authority, under accession (deposition) No. KCTC 0687BP on
10 Nov. 3, 1999.

Example 2: Production of exopolysaccharides employing a novel microorganism, *Enterobacter* sp. SSYL (KCTC 0687BP)

15 In order to produce exopolysaccharides by fermentation using the novel microorganism isolated in Example 1, *Enterobacter* sp. SSYL was cultured under a condition described below: That is, the medium comprising 60g of carbon source, 6g of malt extract, 6g of yeast extract, and 10g of bactopectone was added into a 2L fermentor (Model MDL 6C, Marubishi Co. Ltd., Japan), where glucose was used as a carbon source. A culture condition employed was 30 °C, 250 rpm, 0.25-1.0vvm of air flow,
20 and approximately one day of culture time.

After fermentation, cells were removed from culture medium by ultracentrifugation (Sorvall RC 26 Plus, Rotor SLA-1500, U.S.A.) at 12,000rpm for 20 minutes to obtain supernatant. To the supernatant, acetone was added in the ratio of 1:2(v/v) and a precipitate containing exopolysaccharide was isolated from the
25 supernatant. The isolated precipitate was redissolved in 1.5-2L of distilled water and then filtered under pressure on a filter paper (pore size: 0.7-1.6 µm). The filtrate was dialyzed in the dialysis membrane with 6,000-8,000 MWCO against distilled water for approximately one day. Subsequently, dialyzed solution was freeze-dried to obtain

white dried exopolysaccharide. The final yield of exopolysaccharide against carbon source was 10% or so.

The process for preparing exopolysaccharide by fermentation of novel microorganism described above is outlined in Figure 2. Figure 3 is a graph showing a profile of batch fermentation using glucose as a carbon source.

Example 3: Chemical analysis of exopolysaccharide

Example 3-1: Molecular weight determination of exopolysaccharide

The molecular weight of exopolysaccharide was determined by HPLC with Tosohaas TSK-Gel G-3000SW gel filtration column (7.5mm in diameter, 600mm in length, Japan) and 0.2M ammonium acetate containing 20mM phosphate (pH 6.8) as a mobile phase at a flow rate of 10ml/min. As the size markers for gel filtration chromatography, apoferritin (443kDa), β -amylase (200kDa), alcohol dehydrogenase (150kDa), albumin (66kDa), carboxylate dehydrogenase (29kDa), cytochrome C (12.4kDa), and dextran (2,000Da) were used. After determination of void volume (hereinafter referred to as 'Vo') by passing dextran through the column and elution volume (hereinafter referred to as 'Ve') by passing standard materials (6 kinds of materials used as size markers except dextran) through the column, a standard curve of molecular weight vs. V_e/V_o was prepared. Following the measurement of V_e of exopolysaccharide, V_e/V_o was calculated to determine molecular weight of exopolysaccharide, thus, it has been determined that the exopolysaccharide is a macromolecule with a molecular weight range of 100-1,000kDa.

Example 3-2: Determination of total sugar content of the exopolysaccharide

Total sugar content of the exopolysaccharide was determined by phenol-sulfuric acid reaction. That is, 0.2ml of 5%(w/v) phenol in distilled water was added to 0.2ml of 50 μ g/ml exopolysaccharide in distilled water and then the mixture was shaken, to which 1.0ml of 98% sulfuric acid was added, followed by incubating for 10

minutes, shaking, incubating for 30 minutes, and then sugar content was determined by measuring absorbance at 485nm. A standard concentration curve was prepared using glucose solution (5, 10, 50, 100, and 500 µg of glucose was dissolved respectively, per ml of distilled water) as the standard sugar solution, and the sugar content of exopolysaccharide was determined as $56.9 \pm 19.7\%$. As a reference, when sucrose was used as a carbon source, the sugar content of exopolysaccharide was $52.9 \pm 2.26\%$.

Example 3-3: Analysis of compositions and contents of sugar in the exopolysaccharide

To analyze sugar compositions and their contents of the exopolysaccharides, 8mg of exopolysaccharide treated with 0.8ml of 2M HCl was hydrolyzed at 100 °C for 2-5 hours, and then the reaction mixture was neutralized with barium carbonate, followed by centrifugation to obtain a supernatant, which was subject to HPLC analyses. HPLC analysis was performed with an preparatory column (Waters Carbohydrate column, 4.6mm in diameter, 250mm in length, U.S.A.) and an aqueous solution of 75%(v/v) acetonitrile as a mobile phase at a rate of 0.5ml/min. After 0.5M NaOH solution was run through the column, an electrochemical detection was carried out. Standard concentration curves of monosaccharides were prepared using standard solutions of glucuronic acid, rhamnose, arabinose, glucose, galactose, fucose, and fructose in distilled water with concentration range of 0.2-10mg/ml, respectively.

After all, sugar compositions and their contents of the exopolysaccharide were identified to be 46.7% glucuronic acid, 10.8% fucose, 0.2% fructose, 29.9% glucose, 11.0% galactose and about 1.3% unidentified materials. Sugar compositions of the exopolysaccharide produced by *Enterobacter* sp. reported in the literatures were compared with those produced by the novel microorganism of the invention and the results are summarized in Table 1 below. As shown in Table 1, the sugar compositions of the exopolysaccharides produced by the novel microorganism isolated from root bark of Chinese elm have been demonstrated to be different from those produced by the microorganisms reported in literatures.

Table 1: Comparison of sugar compositions of the exopolysaccharides produced by various

Enterobacter sp.

Microorganism	Sugar Compositions of the Exopolysaccharides
<i>Enterobacter</i> sp.	Glucose, mannose, rhamnose, fucose, glucuronic acid, galacturonic acid
A. <u><i>E. sakazakii</i></u>	Glucose, galactose, fructose, glucuronic acid, acetic acid
B. <u><i>E. agglomerans</i></u>	Glucose, galactose
C. <u><i>E. cloacae</i></u>	Glucose, galactose, fucose, glucuronic acid, pyruvic acid, acetic acid
<i>Enterobacter</i> sp.	Glucose, fucose, glucuronic acid
<i>Enterobacter</i> sp. SSYL(KCTC 0687BP)	Glucose, fructose, fucose, galactose, glucuronic acid

5 Example 3-4: Determination of acidic sugar content of the exopolysaccharide

Acidic sugar content of the exopolysaccharide was determined with the analysis for uronic acid by carbazole method. That is, 50 µg of exopolysaccharide was mixed with 1.5ml of NaBH₄/c.H₂SO₄ (100ml of 98% sulfuric acid containing 0.9g of NaBH₄) on an ice bath with slow shaking followed by vigorous shaking, and then the reaction was continued for 10 minutes in boiling water. After cooling to room temperature, 50 µl of carbazole/anhydrous ethanol (10ml of anhydrous ethanol containing 10mg of carbazole) was added to the reaction mixture, which was, then, heated again in boiling water for 15 minutes, cooled down and analyzed spectroscopically at 525nm. Glucuronic acid standard curve was prepared using glucuronic acid solutions at concentrations of 15, 30, and 50 µg/ml in distilled water, respectively, and acidic sugar content of the exopolysaccharide was calculated to be 9.26±3.03%. As a reference, when sucrose was used as a carbon source, acidic sugar content of exopolysaccharide was 11.8±0.85%.

Example 3-5: Determination of protein content of the exopolysaccharide

Protein content of the exopolysaccharide was determined by the Lowry method with minor modifications: That is, 0.2ml of 0.85%(w/v) NaCl solution containing 500 µg/ml of exopolysaccharide was mixed with 2.2ml of Biuret reagent, to which 0.1ml of Folin-Ciocalteu's reagent was added. After incubation for 30 minutes at room temperature, the reaction mixture was analyzed spectroscopically at 725nm. Bovine serum albumin standard curve was prepared using BSA solutions at concentrations of 250, 500, 750, and 1000 µg/ml in 0.85%(w/v) NaCl solution, respectively, and the protein content of the exopolysaccharide was calculated to be 19.9-0.92%. As a reference, when sucrose was used as a carbon source, protein content of exopolysaccharide was 16.9±2.05%.

The results above, sugar content, acidic sugar content, and protein content of the exopolysaccharide are shown in Figure 4.

Example 3-6: Changes in intrinsic viscosity of the exopolysaccharide depending on concentration and pH, and comparison of rheological properties of exopolysaccharide with those of xanthan gum

The intrinsic viscosities of the exopolysaccharide produced by using sucrose as a carbon source were measured at different concentrations with Hakke viscometer(see: Table 2). The spindle used was S51 and the temperature was 25 °C.

Table 2: Intrinsic viscosity depending on the concentration of the exopolysaccharide

Concentration of the Exopolysaccharide (mg/ml)	Viscosity(cP)
1.1	4.4
9.5	260
15.4	9200

To investigate the pH dependency of the intrinsic viscosity, acid or base was added to the 0.5%(w/v) exopolysaccharide solution and the viscosities were measured (see: Table 3). Initial pH of the 0.5%(w/v) exopolysaccharide solution was 4.1 which belongs to acidic range.

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Table 3: pH dependency of the viscosity

PH	Viscosity(cP)
2.68	76.5
4.10	79.6
7.52	63.8
11.5	50.3

As clearly demonstrated in Table 3 above, the viscosity of exopolysaccharide reached its maximum level at around pH 4, and then decreased gradually with increasing pH. Accordingly, there is a considerable discrepancy between such result of the present invention and the previous report(see: J. of Ferment. Bioeng., 84:13-18, 1997) which showed the viscosity of exopolysaccharide produced by *Enterobacter* sp. gradually increased with increasing pH up to 12, above which it decreased abruptly due to hydrolysis of polysaccharide.

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In order to investigate rheological properties of the exopolysaccharide, 1% exopolysaccharide solution and 1% xanthan gum solution were analyzed for the changes in viscosity and shear stress depending on shear rate, whose results are summarized in Figure 5. As shown in Figure 5, the exopolysaccharide has shown similar pseudoplastic rheological properties to commercially available xanthan gum, moreover, there is almost no difference in steady viscosity between the exopolysaccharide of the invention and xanthan gum.

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Example 4: Determination of immunostimulating activity of the exopolysaccharide in vitro

Anticancer activities of polysaccharides are not attributable to direct cytotoxicity of tumor cell but rather to indirect cure effect by immunostimulation of the body. In order to measure immunostimulating activity of the exopolysaccharide produced in Example 2, following 3 immunological activities were analyzed.

Example 4-1: Immune cell proliferation assay

Following isolation of immune cells from a spleen of Specific pathogen free (SPF) Balb/c mouse, 90 μ l aliquots of immune cell suspension were plated into each well of 96-well microtiter plate to a final concentration of 2×10^6 cells/ml, and 10 μ l each of exopolysaccharide (EPS) solutions were added to the wells to final concentrations of 0.01, 0.1, 1, 10, 100, and 200 μ g/ml, respectively. Immune cell growth was determined by incubation of the 96-well plate at 37 °C for 3 days under 5% CO₂, followed by measuring absorbancy at 490nm within 4 hours after adding 20 μ l aliquots of MTS solution (see: Figure 6). As a positive control, lipopolysaccharide (LPS) solution at the same concentration range mentioned above was used.

Example 4-2: Mixed Lymphocyte Reaction (MLR)

Using spleen cells (5×10^6 cells/ml) from SPF B6C3F1(H-2k) mouse and BDF1(H-2d) mouse, mixed lymphocyte reaction (MLR) were induced by addition of the exopolysaccharide of the invention to the final concentrations of 1, 10, and 100 μ g/ml, followed by incubation at 37 °C for 3 days under 5% CO₂, and then, ³H-thymidine absorbed by the cells was counted (see: Figure 7). As shown in Figure 7, activity of mixed lymphocytes was increased significantly with EPS treatment compared with control (PBS) group.

Example 4-3: Test of direct mitogenicity of immune cells by EPS

200 µl aliquots of spleen cells (1×10^6 cells/ml) from BDF1 mouse were plated into each well of 96-well plate, and the exopolysaccharide solutions were added to the final concentrations of 1, 10, and 100 µg/ml, followed by incubation at 37 °C for 3 days under 5% CO₂, and then, absorption of ³H-thymidine by the cells was counted (see:
5 Figure 8). As shown in Figure 8, the growth rate of immune cells was increased with increasing the concentration of exopolysaccharide, showing that the exopolysaccharide has mitogenic effect on immune cells.

Example 5: Test of acute toxicity of the exopolysaccharide in a septic shock model
10 system

In order to test the acute toxicity of the exopolysaccharide, septic shock induction was evaluated. In this study, 6 week old SPF female ICR mice obtained from the Korea Research Institute of Bioscience and Biotechnology (Taejeon, Korea) were injected peritoneally with exopolysaccharide solutions at doses of 1, 3, 10, 30, and
15 100mg/kg body weight, respectively. Control mice received LPS at dose of 3mg/mouse (150mg/kg) in sterilized distilled water. For experiment, 0.2ml aliquot of EPS or LPS solution per 20g of body weight were injected peritoneally on day 0. Survived mice were counted until day 6 with checking for death 2 times a day. With exopolysaccharide in dose above 10mg/kg, there was a toxic shock symptom similar to
20 septic shock, meanwhile, in dose below 10mg/kg, there was no significant toxicity (see: Figure 9). Based on these results, anticancer activity of exopolysaccharide by virtue of immunostimulation was measured in doses below 3mg/kg, e.g., 0.03, 0.1, 0.3, 1, and 3mg/kg.

Example 6: Evaluation of anticancer activity of the exopolysaccharide in a B16
25 melanoma model system

In order to evaluate anticancer activity of the exopolysaccharide by virtue of immunostimulation, mice implanted with B16 melanoma cells were used for experiments. SPF BDF1 mice (female, 18-22g) were obtained from the Korea Research

Institute of Bioscience and Biotechnology (Taejeon, Korea). The exopolysaccharide solutions were prepared by dilution with sterile distilled water to doses of 0.03, 0.1, 0.3, 1, and 3mg/kg. Solvent used for control group was sterile distilled water, and the positive control mice were injected with adriamycin (ADR) at a dose of 1mg/kg.

5 In this study, ten mice were allotted in each group. On day 0, 0.2ml aliquot of B16 melanoma cell suspension (1×10^5 cells/mouse) was implanted intraperitoneally into each mouse, and 4 hours later, the exopolysaccharide prepared above and solvent were injected peritoneally, respectively. Until day 15, EPS were administered with 16 times in total, and daily dosage was 0.2ml/20g body weight. Consequently, the
10 exopolysaccharide exhibited considerable anticancer activities of 118.5%, 122.2%, 138.1%, 125%, and 112.5% at concentrations of 0.03, 0.1, 0.3, 1, and 3mg/kg, respectively, and among them, 0.3mg/kg group exhibited the highest activity (approximately 140%) (see: Figures 10a, 10b). The positive control agent, adriamycin, showed high survival rate of 177.2%.

15 In order to evaluate toxicity of the exopolysaccharide, changes in body weight were examined, and survived mice were counted 2 times a day. The significant reduction in body weight was not observed until 14 days after exopolysaccharide treatment (see: Figures 11a, 11b). In solvent control group, death of mice began to be observed on day 15, and on day around 34, all mice in treated groups died except mice
20 in positive control group.

As clearly illustrated and demonstrated above, the present invention provides a novel *Enterobacter* sp. isolated from Chinese elm alive, which produces immunostimulating exopolysaccharides with anticancer activity, a process for preparing
25 the exopolysaccharides by culturing the said microorganism. *In vitro* and *in vivo* assays for immunostimulation revealed that the exopolysaccharide of the invention exhibits anticancer activity by virtue of immunostimulation upon tumors, inflammation, edema, digestive system cancer, leukemia, lymphoma, metastatic cancer, and hepatoma. Therefore, the exopolysaccharide of the invention can be used as an active ingredient of

anticancer agents, and as an effective ingredient of immunostimulants for treatment of diseases caused by immune deficiency such as incurable diseases in clinical immunology, cancers, diabetes, male infertility, aquired immunodeficiency syndrome (AIDS), pathogenic viral diseases, and opportunistic diseases, and foodstuffs as well.

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